

**AMYGDALOHIPPOCAMPAL AREA IN ADULT MALE RATS AFTER PROGESTERONE TREATMENT**

LOZANČE OLIVERA\*, MALOBABIĆ S\*\*, ĐELIĆ DIJANA\* and DREKIĆ D\*

\* Faculty of Veterinary Medicine, Belgrade

\*\* Faculty of Medicine, Belgrade

(Received 12. April 2005)

*Neurons of amygdalohippocampal area (AHA) of the amygdaloid complex (AC) in adult control and progesterone (P) neonatally treated adult male rats were studied using light microscopy and morphometric analysis. Progesterone in a single dose of 1.25 mg was administered at the age of 5 days (neonatal period) and in a dose of 5 mg at 30 days of age. Two stereological parameters were estimated: the volume density ( $V_v$ ) ( $mm^0$ ) AHA cell nuclei, cytoplasm, neuropil and the numerical density ( $N_v$ ) ( $mm^{-3}$ ) of neurons. In adult, neonatally and juvenily treated male rats ( $V_v$ ) ( $mm^0$ ) AHA cell nuclei, cytoplasm and numerical density ( $N_v$ ) ( $mm^{-3}$ ) of neurons were significantly higher ( $p < 0.05$  and  $p < 0.001$ ) while  $V_v$  of neuropil was significantly lower ( $p < 0.001$ ) compared with the values of investigated parameters in the controls.*

*Key words: amygdaloid complex, amygdalohippocampal area, progesterone, rats, stereology*

**INTRODUCTION**

Throughout the life span, the brain continues to be shaped and modified by the external world acting through the release and actions of circulating hormones and endogenous growth factors and neurotransmitters (McEwen, 2001). Gonadal steroid hormones are involved in the organization of neural circuits of the developing rat brain with the most dramatic structural changes in the central nervous system occurring during gestation and early postnatal period of life (Arnold and Breedlove, 1985; Arnold and Jordan, 1988; Pilgrim and Hutchinson, 1994; Gorsky, 2000; Merke *et al.*, 2003). They influence the activity and plasticity of neurons and glial cells during early development and they continue to exert trophic and protective effects in the adult nervous system. Steroids, including progesterone (P) produced by gonads and adrenal glands reach the brain, spinal cord and peripheral nerves via the blood stream (Schumacher *et al.*, 2000).

Sex steroid hormones have organizational and activational effects on the sex steroid-accumulating neuronal structures in the central nervous system such as hypothalamic nuclei and nuclei of the amygdaloid complex (AC) (Matsumoto,

1991; Gomes and Newman, 1991; Wood and Coolen, 1997). In the brain regions which have target neurons for sex steroids, neuronal migration, cell death, differentiation of cell types and inhibition or expression of some of neuropeptides involved in the regulation of reproductive functions, are processes that could be modulated by hormones and result in sex differences in neural development (Arnold and Gorsky, 1984; Myceovich *et al.*, 1994; Bradley *et al.*, 1999; Mc Ewen, 2001). The organizational effect of sex steroid hormones also implies permanent influences on a variety of morphological characteristics such as neuronal size, neuron number, dendritic branching, spine density and the synaptic connectivity within different brain areas and different species (Matsumoto, 1991; Gould *et al.*, 1991; Wooly and Mc Ewen, 1993; McEwen *et al.*, 1999). New investigations showed that gonadal steroid hormones play an important role in the proliferation, survival, activation and neuroprotection of neurons. In such a way, Flower *et al.* (2003) showed that newly proliferated cells in the adult male amygdala were affected by gonadal steroid hormones respectively, that gonadal steroid hormones influenced the number of newly proliferated cells in the amygdala. There are newly discovered ways in which gonadal steroid hormones (estrogenic compounds) protect nerve cells from damage, free radicals and neurodegeneration (Lee and McEwen, 2001).

In this work, we investigated the amygdalohippocampal area (AHA), the most caudal part of the sexually dimorphic MN of AC, which poses gonadal steroid receptors (Simerly, 1993, De Vries and Simerly, 2002) and it is very important for the neural control of reproductive behavior (Nishizuka *et al.*, 1981; Mizukama *et al.*, 1983; Gomez and Newman, 1992; Wood and Coolen, 1997). This posterior area of MN including AHA belongs to the medial sexually dimorphic area (SDA) (De Vries *et al.*, 1988, De Vries and Simerly, 2002) and is part of an interconnected vomeronasal pathway and telencephalo - hypothalamic circuit the development of which is influenced by gonadal hormones during early postnatal period (Myceovich *et al.*, 1994; De Vries and Simerly, 2002). Our attention was focused on the long term effects of a single dose of P administered to neonatal and juvenile male rats which were sacrificed in the adult period.

#### MATERIAL AND METHODS

To investigate the influence of P on the morphology of the AHA in adult rat brain after postnatal P treatment, we used two groups of 10 male Wistar rats. The animals were treated: on 5<sup>th</sup> day of life (neonatal period) with 1.25 mg P and on the 30<sup>th</sup> day of life (juvenile period) with 5 mg P. As controls served 10 non-treated age-matched male rats. The animals were sacrificed on the 62<sup>nd</sup> day of life, in the adult period, under ether narcosis. The brains were removed immediately and AC with surrounding structures including anterior hippocampal area were isolated and fixed in Bouin solution. After standard paraffin embedding, serial sections (5  $\mu$ m thick) were stained with hematoxylin-eosin and Herlant methods. Stereological analysis was done by light microscopy (magnification -1088 x).

For stereological analyses, two morphological parameters, the volume density ( $V_v$ ) ( $\text{mm}^0$ ) of AHA neuronal nuclei, cytoplasm, neuropil and numerical density ( $N_v$ ) ( $\text{mm}^{-3}$ ) of neurons were estimated, using the Weibel multipurpose test system (P:42). Sampling was performed so that the most caudal sections in the area of MN were used and 50 test fields were chosen by the intermittent sampling (Kališnik, 1985). The statistical significance was tested with Student's t-test for levels of significance  $p < 0.05$ ,  $p < 0.01$  and  $p < 0.001$ .

## RESULTS AND DISCUSSION

In the adult rat, the amygdalohippocampal area appears immediately caudal to the main body of medial nucleus (MN) and ventral to posterodorsal part of the same nuclei. It consists of well stained, tightly-packed cells, very similar to neurons in the posterior part of MN. On its lateral side this neuronal structure merges with the basomedial nucleus and the ventral part of the lateral entorhinal area. Caudally, AHA is continuous to the hippocampal formation.

The results of stereological analysis of AHA in treated and control male rats are summarized in Tables 1, 2 and 3. Values of investigated stereological parameters i.e. volume density ( $V_v$ ) in control male rats was: 0.052 ( $\text{mm}^0$ ) or 5.2 % for neuronal nuclei, 0.037 ( $\text{mm}^0$ ) or 3.7% for cytoplasm and 0.9104 ( $\text{mm}^0$ ) or 91.04 % for neuropil (Table 1). In neonatally treated adult animals this parameter was: 0.072 ( $\text{mm}^0$ ) or 7.2 % for neuronal nuclei, 0.053 ( $\text{mm}^0$ ) or 5.3% for cytoplasm and 0.8742 ( $\text{mm}^0$ ) or 87.42% for neuropil (Table 2). In juvenily treated adult male rats volume density for neuronal nuclei was 0.064 ( $\text{mm}^0$ ) or 6.4 %, for cytoplasm 0.0619 ( $\text{mm}^0$ ) or 6.19 % and for neuropil 0.8780 ( $\text{mm}^0$ ) or 87.80% (Table 3). Numerical densities ( $N_v$ ) of AHA neurons were:  $10.55 \times 10^4$  ( $\text{mm}^{-3}$ ) in controls,  $17.01 \times 10^4$  ( $\text{mm}^{-3}$ ) in neonatally treated and  $14.05 \times 10^4$  ( $\text{mm}^{-3}$ ) in juvenily treated adult male rats. These results demonstrated that in neonatally and juvenily treated adult male rats ( $V_v$ ) ( $\text{mm}^0$ ) of AHA cell nuclei and of cytoplasm were significantly higher ( $p < 0.05$ ) than in controls, while those of neuropil were significantly lower ( $p < 0.001$ ) than in controls. The numerical densities ( $N_v$ ) ( $\text{mm}^{-3}$ ) of neurons in neonatally and juvenily treated adult male rats were also significantly higher ( $p < 0.001$ ).

Table 1. Investigated stereological parameters of AHA in control male rats

Controls	$\Sigma x$	$\bar{x}$	$\pm SD$	$V_v$ ( $\text{mm}^0$ )	$V_v$ (%)	$N_v$ ( $\text{mm}^{-3}$ )
Nucleus	110	2.20	1.01	0.052	5.2	
Cytoplasm	78	1.56	1.21	0.037	3.7	
Neuropil	1912	38.24	1.01	0.9104	91.04	
N° of neurons	2924	58.48	4.05			$10.55 \times 10^4$

Table 2. Investigated stereological parameters of AHA in neonatally treated adult male rats

Neonatally treated	$\Sigma x$	$\bar{x}$	$\pm SD$	$V_v$ (mm <sup>0</sup> )	$V_v$ (%)	$N_v$ (mm <sup>-3</sup> )
Nucleus	152	3.04	1.18	*0.072	*7.2	
Cytoplasm	96	1.92	1.11	*0.053	*5.33	
Neuropil	1836	36.72	1.71	***0.8742	***87.42	
N <sup>o</sup> of neurons	4378	87.56	3.50			***17.01x10 <sup>4</sup>

Table 3. Investigated stereological parameters of AHA in juvenily treated adult male rats

Juvenily treated	$\Sigma x$	$\bar{x}$	$\pm SD$	$V_v$ (mm <sup>0</sup> )	$V_v$ (%)	$N_v$ (mm <sup>-3</sup> )
Nucleus	136	2.72	1.17	*0.0640	*6.40	
Cytoplasm	67	1.34	0.90	*0.0619	* 6.19	
Neuropil	1916	38.31	1.39	***0.8780	***87.80	
N <sup>o</sup> of neurons	3614	72.28	3.85			***14.05x10 <sup>4</sup>

$\Sigma x$  - total number of intersections;  $\bar{x}$  - mean value;  
SD - standard deviation;  $V_v$  - volume density;  $N_v$  - numerical density.  
\*-p<0.05      \*\*-p<0.01      \*\*\*-p<0.001

In the present report we investigated the influence of P on AHA as part of posterior AC in adult rats brain which were treated during postnatal period of life (in the neonatal and in the juvenile period). Perinatal development is often viewed as the major period of time for organizing of steroid – sensitive neural circuits by steroid hormones. Behavioral and neuroendocrine responses to steroids are dramatically different before and after puberty, suggesting that puberty is another period of time during which gonadal steroids affect neural development (Romeo *et al.*, 2000). Peripheral steroid hormones act on brain tissues through intracellular receptor-mediated mechanisms to regulate several important brain neuronal functions (Schumacher *et al.*, 2000). Most hormonal actions in the brain are mediated by their cognitive nuclear receptors which were ligand-regulated transcription factors (Tsai and OMalley, 1994). In different brain areas with sex steroid accumulating neuronal structures (such as AC), sex steroids promote neuronal and glial differentiation (Mong *et al.*, 1999) and induce permanent sexual dimorphism: in nuclear volume (Gorsky *et al.*, 1978), neuronal number (Guillamon *et al.*, 1998; Lozanče *et al.*, 1994;1995), distribution patterns of serotenergic and vasopressinergic fibers (Simerly *et al.*, 1982; De Vries *et al.*, 1983), synaptic formation and neuronal connectivity (Nishizuka and Arai, 1981; Sakuma and Pfaff, 1981) and in soma size of neurons (Bradley *et al.*, 1999). The effects od steroids in

the modulatory role of the AC may be expressed via the involvement of steroid sensitive neurons and the presence of highly specific steroid receptors (Yokosuka *et al.*, 1997; Nishikara *et al.*, 2003) or via rapid regulatory effects on several membrane-associated and intracellular responses that do not depend on changes in gene expression (nongenomic mechanisms) (Beyer and Karolczak, 2000; McEwen 2001). These include changes in the electrical properties of neurons, as well as alterations in neurotransmitter release (Navarro *et al.*, 2003). Some steroid hormones, named "neurosteroids" can also be synthesized within the nervous system. They include progesterone, P and their reduced metabolites and sulfate esters (Akawa and Baulieu, 1999; Schumacher *et al.*, 2000; Mayo *et al.*, 2001; Semeniuk *et al.*, 2001).

Generally, present stereological analysis showed that treatment with P in neonatal and juvenile (late puberty) periods of life caused effects on AHA in male rats. That is demonstrated by the values of investigated morphometric parameters in adult rats. The value of  $N_v$  in neonatally and juvenily treated male rats showed a significant increase in the number of neurons and in volume density of AHA neuronal nuclei and cytoplasm (in regard to values in control male rats). Beyer and Feder (1987) described that perinatal gonadal steroids produced observable changes in behavior and physiology that were not concurrent with the first administration of the hormone but manifested themselves several weeks or months after a second application of the hormonal stimulus. In the brain, most hormonal actions are mediated by their cognate nuclear receptors which are ligand-regulated transcription factors (Tsai and O'Malley, 1994). Đelić *et al.* (2003) in adult male rats brain, after neonatal P treatment, observed the long term effects on the number of neurons in different nuclei of AC, as well as in other brain nuclei. Perinatal steroids probably produced immediate cellular changes that were not manifested until much later, when appropriate hormonal conditions were present (production of adequate amounts for sex steroids beginning puberty). Siburg *et al.* (1991) suggested that the plasticity of sex steroids receptor system in the rodent brain is not only dependent on development, but also on hormonal status of the animals. In the moment of sacrifice, the animals in our experiment were in the adult period when adequate hormonal conditions have been present, as well as in juvenile treated animals. Possible genomic effects, consisted of a shortening of latency and an enhancement of the response to a second administration of the hormone. At the cellular level, this change in responsiveness is apparently related to a persistent change in chromatin structure that allows the receptor-hormone complex to interact more efficiently with specific sites on the DNA (Burch and Wientraub, 1983). Progesterone presumably binds to the receptor complex which then stimulates the protein synthesis mechanism of the cell. Increases in specific cytoplasmic organelles including rough endoplasmic reticulum, polyribosomes and proliferation of Golgi apparatus after treatment with sex steroid hormones, suggest a stimulation of protein synthetic capacity of the cell (Carrer and Aoki, 1982; Meisel and Pfaff, 1985). It has been shown that P as "neurosteroid" plays an important role in the formation of new myelin sheath and that it promotes myelination by activating the gene expression of genes coding myelin proteins (Schumacher *et al.*, 2000). Progesterone, as a neurosteroid in Purkinje neurons

only during neonatal life, may be involved in the promotion of neuronal and glial growth and neuronal synaptic contact in the cerebellum (Tsutsui *et al.*, 2000). Rupprecht (1997) demonstrated that some of steroid hormones including derivatives of P as neuroactive steroids can regulate also gene expression via the P receptor after intracellular oxidation. He described that in physiological concentrations these neuroactive steroids regulated the neuronal function through their concurrent influence on transmitter-gated ion channels and gene expression. Neurons of AHA of AC through receptors and genomic expression may also be stimulated to increase a production of cytoskeleton elements or membranous components that contribute to the maintenance of their structure. However, as in our earlier studies, we have not ruled out the possibility that the observed changes in AHA are secondary steroid-mediated changes provoked elsewhere in the brain, which in turn may result in altered inputs to neuronal structures of AC.

Address for correspondence:  
Olivera Lozanče, PhD  
Faculty of Veterinary Medicine  
Department of Anatomy  
Bul oslobođenja 18, Belgrade  
Serbia & Montenegro  
e-mail: dijana@vet.bg.ac.yu

#### REFERENCES

1. Akawa YEE, 1999, Neurosteroids: behavioral aspects and physiological implications, *J Soc Biol*, 193,3, 293-8.
2. Arnold AP, Jordan CL, 1988, Hormonal organization in neural circuits. *In: Frontiers in Neuroendocrinology*, 10, Raven Press, New York.
3. Arnold A, Gorski R, 1984, Gonadal steroid induction of structural sex differences in the central nervous system, *Annu Rev Neurosci*, 7, 413-42.
4. Arnold A, Breedlove S. 1985, Organizational and activational effects of sex steroids on the brain and behavior: a re-analysis. *Horm Behav*, 19, 469-98.
5. Beyer C, Feder HH, 1987, Sex steroids and afferent input: Their roles in brain sexual differentiation, *Ann Rev Physiol*, 49, 349-64.
6. Beyer C, Karolczak M, 2000, Estrogenic stimulation of neurone growth in midbrain dopaminergic neurons depends on cAMP/protein kinase A signaling, *J Neurosci Res*, 59, 1, 107-16.
7. Bradley M, Breedlove SM, 1999, A brain sexual dimorphism controlled by adult circulating androgens, *Proc Nat Acad Sci USA*, 96, 7538-40.
8. Burch JBE, Wientraub H, 1983, Temporal order of chromatin structural changes associated with activation of the major chicken vitellogenin gene, *Cell*, 33, 65-76.
9. Canteras NS, Simerly RB, Swanson LW, 1995, Organization of projections from the medial nucleus of the amygdala: a PHAL study in the rat, *J Comp Neurol*, 360, 2, 213-45.
10. Carrer HF, Aoki A, 1982, Ultrastructural changes in the hypothalamic ventromedial nucleus of ovariectomized rats after estrogen treatment, *Brain Res*, 240, 221-33.
11. Cherry JA, Basham ME, Weawer CE, Krohmer RW, Baum MJ, 1990, Ontogeny of the sexually dimorphic male nucleus in the preoptic / anterior hypothalamus of ferrets and its manipulation by gonadal steroids, *J Neurobiol*, 21, 6, 844-57.
12. Flower CD, Freeman ME, Wang Z, 2003, Newly proliferated cells in the adult male amygdala are affected by gonadal steroid hormones, *J Neurobiol*, 57, 3, 257-69.

13. DeVries GJ, Best W, Sluiter AA, 1983, The influence of androgens on the development of a sex difference in the vasopressinergic innervation of the rat lateral septum, *Dev Brain Res*, 8, 377-408.
14. De Vries GJ, Gonzales CL, Yahr P, 1988, Afferent connections of the sexually dimorphic area of the hypothalamus of male and female gerbils, *J Comp Neurol*, 271,1, 91-105.
15. De Vries GJ, Simerly RB, 2002, Anatomy, Development and Function of Sexually Dimorphic Neural Circuits in the Mammalian Brain, *Horm Brain Behavior*, 64, 4, 137-83.
16. Đelić Dijana, Lozanče Olivera, Nikolić Zora, Blagojević Zdenka, Mrvić-Jović Verica et al. 2003, Changes in myelination of neurons in different brain regions in progesterone-treated rats, *Acta Veterinaria*, 53, 5-6, 367-75.
17. Gomez DM, Newman SW, 1991, Medial nucleus of the amygdala in the adult Syrian hamster: A Quantitative Golgi analysis of gonadal hormonal regulation of neuronal morphology, *Anat Rec*, 231, 4, 498-509.
18. Gomez DM, Newman SW, 1992, Differential projections of the anterior and posterior regions of the medial amygdaloid nucleus in the Syrian hamster, *J Comp Neurol*, 317, 2, 195-218.
19. Gorsky RA, Gordon JH, Shyrne JE, Sautham AM, 1978, Evidence for a morphological sex difference within the medial preoptic area of the rat brain, *Brain Res*, 148, 333-46.
20. Gorsky RA, 2000, Sexual differentiation of the nervous system, In: Principles of neural science, 1131-46, New York: McGraw-Hill.
21. Gould E, Woolly SC, McEwen B, 1991, The hippocampal formation: morphological changes induced by thyroid, gonadal and adrenal hormones, *Psychoneuroendocrinol*, 16, 1-3, 67-84.
22. Guillamon A, Segovia S, Del Abril A, 1988, Early effects of gonadal steroids on the neuron number in the medial posterior region and the lateral division of the bed nucleus of the stria terminalis in the rat, *Dev Brain Res*, 44, 281-90.
23. Kališnik M, 1985, Temelji stereologije, *Stereol Jugosl, suppl.* II, 3, 1-143.
24. Lee SJ, McEwen BS, 2001, Neurotropic and neuroprotective action of estrogens and their therapeutic implications, *Ann Rev Pharmacol Toxicol*, 41, 569-91.
25. Lozanče Olivera, Drekić D, Malobabić S, Cvetković Dijana, 1994, Sex differences in posterodorsal subregion of the medial nucleus (MN) of the amygdaloid complex (AC) in the rat brain, Stereological analysis, *Folia Anatomica*, 21-22, suppl 1, *Book of Abstracts of XXIII Congress of YAA with International Participants*, 22.
26. Lozanče Olivera, Drekić D, Malobabić S, Cvetković Dijana, 1995, The study of sex differences in anterior and posterodorsal subregions of the medial nucleus (MN) of the amygdaloid complex (AC) in the rat brain, *Acta Veterinaria (Beograd)*, 45, 2-3, 103-8.
27. Matsumoto A, 1991, Synaptogenic action of sex steroids- in developing and adult neuroendocrine brain, *Psychoneuroendocrinol*, 16, 1-3, 25-40.
28. Mayo W, LeMoal M, Abrous DN, 2001, Pregnenolone sulfate and aging of cognitive functions: behavioral, neurochemical and morphological investigations, *Horm Behav*, 40, 2, 215-7.
29. McEwen B, 2001, Genome and hormones: Gender differences in Physiology. Invited review: Estrogen effects on the brain: multiple sites and molecular mechanisms, *J Appl Physiol*, 91, 2785-801.
30. McEwen BS, Tanapat P, Weiland NG, 1999, Inhibition of dendritic spine induction on hippocampal CA1 pyramidal neurons by a non-steroidal estrogen antagonist in female rats, *Endocrinol*, 140, 1044-7.
31. Meisel RL, Pfaff DW, 1985, Brain region specificity in estradiol effects on neuronal ultrastructure in rats, *Mol Cell Endocrinol*, 40, 159-66.
32. Merke PD, Fields JD, Keil MF, Vaituzis CA, Chrousos, GP et al., 2003, Children with classic congenital adrenal hyperplasia have decreased amygdala volume: potential prenatal and postnatal hormonal effects, *J Clin Endocrin Metabolism*, 88, 4, 1760-5.
33. Mizukami S, Nishizuka M, Arai Y, 1983, Sexual difference in nuclear volume and its ontogeny in the rat amygdala, *Exp Neurol*, 79, 569-75.

34. Mong Jessica A, Glaser E, McCarthy Margaret M, 1999, Gonadal steroids promote glial differentiation and alter neuronal morphology in the developing hypothalamus in a regionally specific manner, *J Neurosci*, 19,4, 1464-72.
35. Myceovich PE, Abelson L, Fok H, Ulibarry C, Priest CA, 1994, Gonadal steroid control of preprocholecystokinin mRNA expression in the limbic-hypothalamic circuit: comparison of adult with neonatal steroid treatment, *J Neurosci Res*, 38, 4, 386-98.
36. Navarro CE, Saeed SA, Murdock Cynthia, Martinez-Fuentes AJ, Arova KK et al., 2003, Regulation cyclic adenosine 3',5'-monophosphate signaling pulsatile neurosecretion by G-i coupled plasma membrane. Estrogen receptors immortalized gonadotropin-releasing hormone neurons, *Mol Endocrin*, 17,9, 1792-804.
37. Nishizuka M, Arai Y, 1981, Sexual dimorphism in synaptic organization in the amygdala and its dependence on neonatal hormone environment, *Brain Res*, 212, 31-8.
38. Nishihara E, Yoshida-Komiya H, Chan CS, Liao L, Davis RL et al., 2003, SRC-1 null mice exhibit moderate motor dysfunction and delayed development of cerebellar Purkinje cells, *J Neurosci*, 23, 1, 213-22.
39. Pilgrim C, Hutchinson JB, 1994, Developmental regulation of sex differences in the brain: can the role of gonadal steroids be redefined? *J Comp Neurosci*, 60, 843-55.
40. Romeo DR, Diedrich SL, Sisk CL, 2000, Effect of gonadal steroids during pubertal development and androgen and estrogen - immunoreactivity in the hypothalamus and amygdala, *J Neurobiol*, 44, 3, 361-8.
41. Rupprecht R, 1997, The neuropsychopharmacological potential of neuroactive steroids, *J Psychiatr Res*, 31, 3, 297-314.
42. Sakuma Y, Pfaff DW, 1981, Electrophysiological determination of projections from ventromedial hypothalamus to midbrain central gray: difference between female and male rats, *Brain Res*, 225, 184-8.
43. Semeniuk T, Jhangri GS, Le Melleo JM, 2001, Neuroactive steroid levels in patients with generalized anxiety disorder, *J Neuropsych Clin Neurosci*, 13 3, 396-8.
44. Siburg RM, Stumpf WE, Shughrue PJ, Hochberg RB, Drews U, 1991, Distribution of estrogen target sites in the day old mouse brain and pituitary gland during the critical period of sexual differentiation, *Develop Brain Res*, 61, 11-22.
45. Simerly RB, Swanson LW, Gorsky RA, 1982, Demonstration of a sexual dimorphism in the distribution of serotonin-immunoreactive fibers in the medial preoptic nucleus of the rat, *J Comp Neurol*, 225, 151-66.
46. Simerly RB, 1993 Distribution and regulation of steroid hormone receptor gene expression in the central nervous system, *Adv Neurol*, 59, 207-26.
47. Schumacher M, Akawa Y, Guennoun R, Robert F, Labombarda F et al., 2000, Steroid synthesis and metabolism in the nervous system: trophic and protective effects, *J Neurocytol*, 29, 5-6, 307-26.
48. Swann J, Fiber JM, 1997, Sex differences in function of a pheromonally stimulated pathways: role of steroids and the main olfactory system, *Brain Res*, 44, 4, 409-13.
49. Tsutsui K, Ukena K, Usui M, Sakamoto H, Takase M, 2000, Novel brain function: biosynthesis and actions of neurosteroids in neurons, *Neurosci Res*, 36, 4, 261-73.
50. Wong M, Moss RL, 1992, Modulation of single - unite activity in the rat medial amygdala by neurotransmitters, estrogen priming, and synaptic inputs from the hypothalamus and midbrain, *Synapse*, 10, 2, 94-102.
51. Wood RI, Coolen LM, 1997, Integration of chemosensory and hormonal inputs is essential for sexual behavior in male Syrian hamster: role of the medial amygdaloid nucleus, *Neurosci*, 78, 4, 1027-35.
52. Woolley CS, McEwen B, 1993, Roles of estradiol and progesterone in regulation of hippocampal dendritic spine density during the estrous cycle in the rat, *J Comp Neurol*, 336, 293-306.
53. Yokosuka M, Okamura H, Hayashi S, 1997, Postnatal development and sex difference in neurons containing estrogen receptor-alfa immunoreactivity in the preoptic brain, the diencephalon and the amygdala in the rat, *J Comp, Neurol*, 389, 1, 81-93.

**AMIGDALO-HIPOKAMPALNA ZONA ODRASLIH MUŽJAKA PACOVA TRETIRANIH  
PROGESTERONOM**

LOZANČE OLIVERA, MALOBABIĆ S, ĐELIĆ DIJANA i DREKIĆ D

SADRŽAJ

Morfometrijskom analizom uz pomoć svetlosne mikroskopije ispitivani su neuroni amigdalo-hipokampalne zone (AHA) amigdaloidnog kompleksa (AK) odraslih mužjaka pacova - kontrola i odraslih mužjaka pacova tretiranih progesteronom. Progesteron je aplikovan u dozama od 1,25 mg 5. dana (u neonatalnom periodu) i 5 mg - 30. dana (u kasnom juvenilnom) periodu života. Praćena su dva stereološka parametra, zapreminska gustina ( $V_v$ ) ( $\text{mm}^0$ ) jedra neurona, citoplazme i međučelijskog prostora, i numerička gustina neurona ( $N_v$ ) ( $\text{mm}^{-3}$ ). U neonatalno i juvenilno tretiranih mužjaka pacova volumenska gustina ( $V_v$ ) ( $\text{mm}^0$ ) AHA jedara neurona i citoplazme i numerička gustina ( $N_v$ ) ( $\text{mm}^{-3}$ ) neurona odraslih mužjaka pacova bile su značajno povećane ( $p < 0,05$  i  $p < 0,001$ ), dok je  $V_v$  neuropila bila značajno niža ( $p < 0,001$ ) u poređenju sa vrednostima ispitivanih parametara kod kontrolnih životinja.